

Anti-Tilapia (*Oreochromis niloticus*) IgM monoclonal antibody

Product no: F04

**Product Information** 



## **Product Description**

This monoclonal antibody (Mab) reacts with Anti-Tilapia (Oreochromis niloticus) immunoglobulin M (IgM). The Mab is of an IgG1 isotype and recognises the heavy chain of the molecule.



# **Use of product**

The Mab is recommended for use in an Enzyme-Linked Immunosorbent Assay (ELISA) for measuring antibody levels of antigen-induced IgM. The optimal conditions for use of this product vary depending on the procedure used. The user must determine the suitability of the product for a particular procedure. This product is for *in vitro* use only.



## Vial Contents

Each vial contains  $200\,\mu$ g of lyophilised protein prepared from bovine-free culture medium and contains no animal-derived stabilisers. This is sufficient for three 96-well ELISA plates.

#### The product should be reconstituted as follows:

Add 1 ml of phosphate buffered saline (PBS) (see buffers) to the vial, and store in aliquots. Dilute  $1_{33}$  in antibody buffer before use.



## Storage

Store at -20°C or below prior to reconstitution. For prolonged storage, the Mab solution should be stored at -20°C as working aliquots. Repeated freeze thawing of the product should be avoided.

## Suggested protocol for the detection of Tilapia IgM by indirect - Enzyme-Linked Immunosorbent Assay (ELISA)

The method outlined here is only a guideline and assay conditions may vary depending on the antigen used to coat the ELISA plate and environmental conditions.

## Procedure

The coating procedure depends on the type of antigen used in the screening.

Plates coated with particulate antigens (eg bacteria)

- Coat 96-well ELISA plate with 0.05% (w/v) poly-L-lysine in coating buffer, 50  $\mu \rm l$  well^1 for 60 min
- Wash plate with 2 washes of low salt wash buffer
- Resuspend bacteria in PBS (1 x10<sup>8</sup> bacteria ml <sup>-1</sup>) and add to the wells of the ELISA plate at 100  $\mu$ l well<sup>-1</sup>. Incubate overnight at 4°C or centrifuge plate at x 200 g for 5 min and incubate for 60 min at  $\approx$ 22°C
- Add 50  $\mu l$  well  $^1$  0.05% (v/v) gluteraldehyde, diluted in PBS, to the antigen and incubate for a further 20 min at  ${\approx}22^{\circ}{\rm C}$

Plates coated with soluble antigen

 Coat 96 well ELISA plate with 100 μl well<sup>-1</sup> antigen [(1-20 μg ml<sup>-1</sup>) this will need to be optimised by the user] dissolved in coating buffer. Cover and incubate overnight at 4°C



## The remainder of procedure is as follows:

- Wash plate 3 times with low salt wash buffer
- Post-coat plate (to block non-specific binding sites) with either 1% (w/v) bovine serum albumin (BSA) or 3% (w/v) casein (dried milk). Add 250 $\mu$ l well<sup>-1</sup> and incubate for 2 h at  $\approx$ 22°C
- Wash plate with 3 washes of low salt wash buffer
- Prepare doubling-dilutions of the fish serum in PBS starting with a 1/2 dilution. Also use doubling-dilutions of pre-immune serum or serum from non-vaccinated/non-diseased fish, and PBS as negative controls. Add serum and control dilutions to the wells (100µl well<sup>-1</sup>) and incubate for 3 h at ≈22°C or overnight at 4°C
- Wash plate with 5 washes of high salt wash buffer, incubating for 5 min on last wash
- Add 100 $\mu$  well  $^{_1}$  of the reconstituted anti-fish Mab and incubate for 60 min at  $\approx\!22^\circ\text{C}$
- Wash plate with 5 washes of high salt wash buffer, incubating for 5 min on last wash
- Add 100 $\mu$ l well<sup>-1</sup> conjugate (anti-mouse IgG-HRP diluted 1/1000 in conjugate buffer). Incubate for 60 min at  $\approx$ 22°C
- Wash plate with 5 washes of high salt wash buffer, incubating for 5 min on last wash
- Add 100 $\mu$ l well^1 chromogen in substrate buffer and incubate for 10 min at  $\approx\!22^\circ\text{C}$
- Stop reaction with  $50\mu$ l well<sup>-1</sup> of stop solution
- Read plate at 450 nm in an ELISA reader. Blank ELISA reader against wells filled with chromogen and stop solution. If an ELISA reader is unavailable compare colour with that of background (i.e. negative control).

## NB. WEAR GLOVES WHEN USING CHROMOGEN AND STOP SOLUTION



# Buffers

## Coating buffer (Carbonate-bicarbonate solution)

Na<sub>2</sub>CO<sub>3</sub> 1.59 g NaHCO<sub>3</sub> 2.93 g **Dissolved in one litre of distilled water. Adjusted to pH 9.6.** N.B. prepare fresh coating buffer on each occasion

## Phosphate Buffered Saline (PBS)

0.02M Phosphate,	0.15M NaCl
NaH,PO,2H,O	0.876g
Na,HPO,2H,O	2.56g
NaĈI	8.77g

Dissolved in one litre of distilled water. Adjusted to pH 7.3 with conc. HCI

## Wash buffer (x10) (low salt)

Trisma base	24.2 g
NaCl	222.2 g
Merthiolate	1 g
Tween 20	5 ml

Dissolved in one litre of distilled water. Adjusted to pH 7.3 with conc. HCI

#### Wash buffer (x10) (high salt)

Trisma base	24.2 g
NaCl	292.2g
Merthiolate 1 g	
Tween 20	10 ml

Dissolved in one litre of distilled water. Adjusted to pH 7.7 with conc. HCI

#### Antibody buffer

Add 1 g of BSA to 100 ml of PBS (i.e. 1 % BSA solution)



<u>Conjugate buffer</u> Add 1g of BSA to 100 ml of low salt wash buffer

## Substrate buffer (Sodium acetate/ citric acid buffer)

Citric acid 21.0 g Sodium acetate 8.2 g Dissolved in one litre of distilled water. Adjusted to pH 5.4 with 1 M NaOH Add 5  $\mu$ l of H<sub>2</sub>O<sub>2</sub> to 15 ml substrate buffer

### **Substrate**

Prepare 3'3'5'5'-Tetramethylbenidine dihydrochloride (TMB) (42 mM) in 1:2 acetic acid: distilled water. Add 150  $\mu$ l of this solution to 15 ml substrate buffer

#### Stop reagent

2M H<sub>2</sub>SO<sub>4</sub> in distilled water



# **Certificate of Analysis**

Anti-Tilapia (Oreochromis niloticus) IgM monoclonal antibody

Product no.

Batch no.

Date of expiry

## Absorbance of reconstituted Mabs by Indirect ELISA

The reconstituted Mab gives an absorbance of at 450nm by ELISA when the plate is coated with  $10\mu$ g/ml purified IgM.



Aquatic Diagnostic Ltd., Institute of Aquaculture, University of Stirling, Stirling, Scotland, FK9 4LA

Telephone: +44 (0)1786 466568 Fax: +44 (0)1786 472133 E-mail: aquaticdiagnostics@stir.ac.uk http://www.aquaticdiagnostics.com