

The logo for AQUATIC Diagnostics Ltd features the word "AQUATIC" in a bold, white, sans-serif font, set against a dark blue rectangular background. Below this, the words "Diagnostics Ltd" are written in a lighter blue, sans-serif font. The entire logo is centered on a light blue background with decorative wavy lines in shades of blue and white.

**AQUATIC**  
Diagnostics Ltd

Anti-Koi Herpesvirus (KHV)  
monoclonal antibody

Product no: P14

Product Information



## Product Description

The monoclonal antibody (Mab) against Koi Herpesvirus (KHV) is specific for this virus. The specificity of the Mab has been tested by IFAT against a range of viral pathogens that infect fish, including infectious pancreatic necrosis virus (IPNV, Birnaviridae), infectious haematopoietic necrosis virus (IHNV, Rhabdoviridae), spring viraemia of carp virus (SVCV, Rhabdoviridae), viral haemorrhagic septicaemia virus (VHSV, Rhabdoviridae), European catfish virus (ECV, Iridoviridae), eel herpesvirus (Herpesviridae), and infectious salmon anaemia virus (ISAV, Orthomyxoviridae). The Mab is of an IgG1 isotype.



## Use of product

The Mab is recommended for use in immunohistochemistry (IHC), but can also be used in IFAT. The optimal conditions for use of this product vary depending on the procedure used. The user must determine the suitability of the product for a particular procedure. This product is for *in vitro* use only.



## Vial Contents

Each vial contains 200  $\mu$ g of lyophilised protein prepared from bovine-free culture medium and contains no animal-derived stabilisers. This is sufficient for between 100-200 tests depending on the area of tissue to be screened in IHC.

The product should be reconstituted as follows:

- Add 1 ml of phosphate buffered saline (PBS) (see buffers) to the vial and store as aliquots. Dilute 1/10 in PBS before use.



## Storage

Store at -20°C prior to reconstitution. For prolonged storage, the Mab solution should be stored at -20°C. Repeated freeze/thawing of the product should be avoided.



## Protocol

### **Suggested protocol for the detection of KHV in fixed tissue sections by immunohistochemistry**

This procedure has been developed to work on tissues fixed in 10 % buffered formalin for 24 hours. Individual protocols may have to be developed depending upon the tissue examined, fixation etc.

### **Procedure**

- .. Prepare paraffin-embedded tissue sections.
- .. Dewax and rehydrate sections in xylene (2 x 5min), 100% ethanol (5 min), 70% ethanol (3 min), then rinse in distilled water.
- .. Place slides in a humid chamber.
- .. Keep sections moist at all times - do not allow them to dry out.
- .. Mark rings around the tissue sections using a wax PAP pen.
- .. Block endogenous peroxidase activity by incubating the slides for 10 min at room temperature ( $\approx 22^{\circ}\text{C}$ ) with  $\text{H}_2\text{O}_2$  in methanol (see buffers).
- .. Wash the slides three times with Tris buffered saline (TBS) (see buffers).

- “ Block non-specific binding sites with normal goat serum diluted 1/10 in TBS for 10 min at room temperature.
- “ Pour off the serum and remove excess serum tapping the slide edges on a paper towel.
- “ Place 50-100  $\mu$ l of reconstituted anti-KHV Mab onto the tissue sections (the volume added will depend on the size of sample to be covered) and incubate for 60 min at room temperature in a humid chamber.
- “ Use appropriate controls i.e. known positive tissue as a positive control and uninfected tissue as a negative control; these should both be incubated with the reconstituted Mab and PBS separately.
- “ Wash slides three times with TBS.
- “ Add goat anti-mouse IgG biotin conjugate (1/100 in TBS) to the slides for 30 min
- “ Wash slides three times with TBS
- “ Add streptavidin-horseradish peroxidase (1/100 in TBS) to the slides for 30 min
- “ Wash slides three times with TBS
- “ To visualise the reaction, incubate the slides for 10 min with DAB solution (see buffers) or with a commercially available True Blue staining kit following the manufactures instructions.
- “ If stained with DAB stop the reaction by immersing the slides in tap water and counter-stain them with haematoxylin for 3-4 min.
- “ Rinse in tap water for 10 min.
- “ Dehydrate the slides in 70% ethanol (3 min), 100% ethanol (5 min), xylene (2 x 5 min)
- “ Mount the slides with Pertex and leave in fume cupboard to set.
- “ Examine tissue under a light microscope –cells infected with the virus appear golden brown in colour when stained with DAB.



## Buffers

### Phosphate buffered saline (PBS)

0.02M Phosphate, 0.15M NaCl                      pH adjusted to 7.2 with HCl

$\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$                       0.876g/l

$\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$                       2.56g/l

NaCl                                      8.77g/l

### Tris buffered saline (TBS)

Trisma base                              2.42g

NaCl                                      29.24g

Dissolve in approximately 900 ml distilled water, adjust pH to 7.2 using HCl and make up to 1 litre

10% (v/v) Hydrogen peroxide in methanol

Add 1ml H<sub>2</sub>O<sub>2</sub> (30% v/v solution) to 9 ml methanol

3,3'-Diaminobenzidinetetrahydrochloride (DAB)

Dissolve one 10mg tablet DAB in 6.67mls TBS

Place 0.5 ml aliquots of the solution into bijoux bottles, store at -20°C.

For use add 5mls TBS and 0.1ml 1 % H<sub>2</sub>O<sub>2</sub> to 0.5 ml aliquot

NB. DAB is a possible carcinogen



## Certificate of Analysis

Anti-Koi Herpesvirus (KHV) monoclonal antibody

Product no. P14

Batch no.

Date of expiry

**Activity in IHC:** Cells infected with the virus appear golden brown in colour when stained with DAB.



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